

Imaging Cores – Electron Negative Staining TEM SOP

Preparation performed by investigator prior to sample submission to the Core Facility

- ⁽¹⁾ Fixation with 2.5 % glutaraldehyde in 0.1M PBS for 1 hr.
- Transfer to 0.1 M PBS buffer pH 7.2 - 7.4
- Submit the suspension to the core facility in the buffer, solution, or double distilled water.

Procedure

Load 5 μ L of ^(2,3,4) sample suspension in a ⁽⁵⁾ carbon-coated grid	⁽⁶⁾ 1min <input type="checkbox"/> 2 min <input type="checkbox"/> 5 min <input type="checkbox"/>
Remove excess with filter paper.	<input type="checkbox"/>
Staining with 5 μ L 2 % UA.	1 min <input type="checkbox"/> 2 min <input type="checkbox"/>
Remove excess with filter paper.	<input type="checkbox"/>
Rinse in 0.1 M PBS <input type="checkbox"/> or ddH ₂ O <input type="checkbox"/>	3 x 10 secs <input type="checkbox"/> <input type="checkbox"/> <input type="checkbox"/>
Rinse in ddH ₂ O.	1 x 10 secs <input type="checkbox"/>
Let the grid dry.	5 min <input type="checkbox"/>

Notes:

1. Fixation is required for bacteria, virus, protozoa microorganisms.
2. Exosomes and liposomes preparation in PBS, nanoliposomes, nanocellulose or proteins can be used for this procedure without prior fixation.
3. Powder (lyophilized) samples require to be resuspended in double distilled water, or specific solution.
4. Prepare several dilutions of the sample in the buffer or molecular grade water accordingly. Include an undiluted, and an unstained sample as well.
5. To increase the sample adhesion to the carbon-coated grids, glow discharge the grids for 30 sec at 15 mA.
6. Time of incubation will depend on the sample and the concentration of the suspension.